

# Improving Chromatographic Performance of Underivatized Anionic Polar Pesticides in Food to Overcome Renowned Analytical Challenges

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Food & Environmental Markets



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## Introduction

Waters

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- Multiresidue analyses determine as many residues as possible in the smallest number of analyses
  - Generic extraction, no/limited cleanup, highly selective determination step (GC- and LC-MS/MS or HRMS)
  - A number of different very successful implementations
    - *e.g.* QuEChERS, mini Luke...
- Polar pesticides in many cases are not amenable to the generic multiresidue approach as they are challenging to analyse.
- The source of these difficulties arise from the physicochemical properties of these compounds, which impact and complicate each stage of the analysis.
- Historically these compounds have been analysed in a series of selective single residue methods (SRM), adding significant costs so were often excluded from surveillance.
- As well as glyphosate, EU screening labs also want to include AMPA (glyphosate metabolite) and a number of other challenging polar pesticides in a single method.

## What and Why?

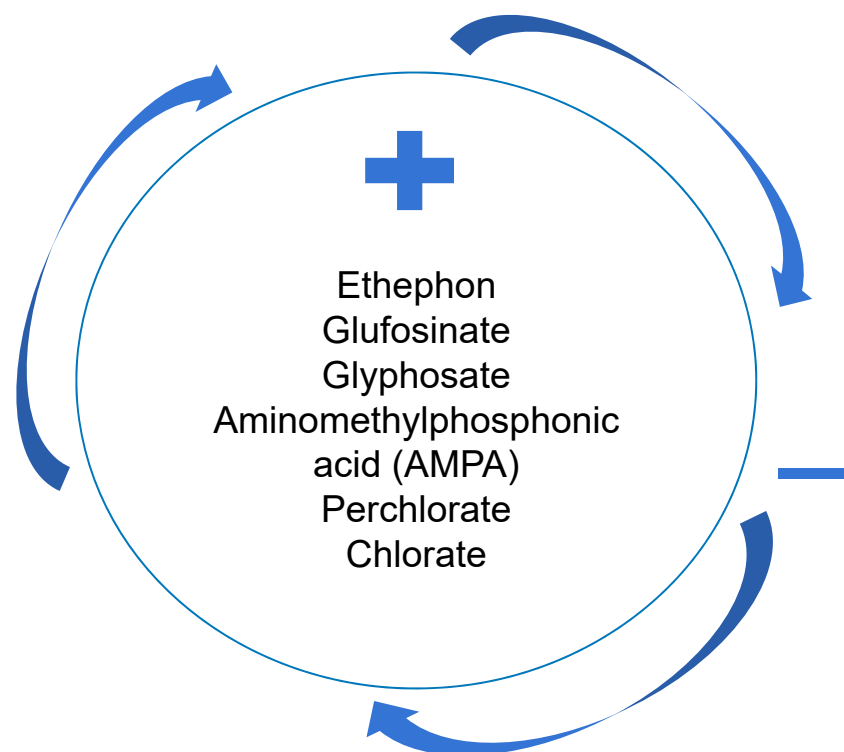
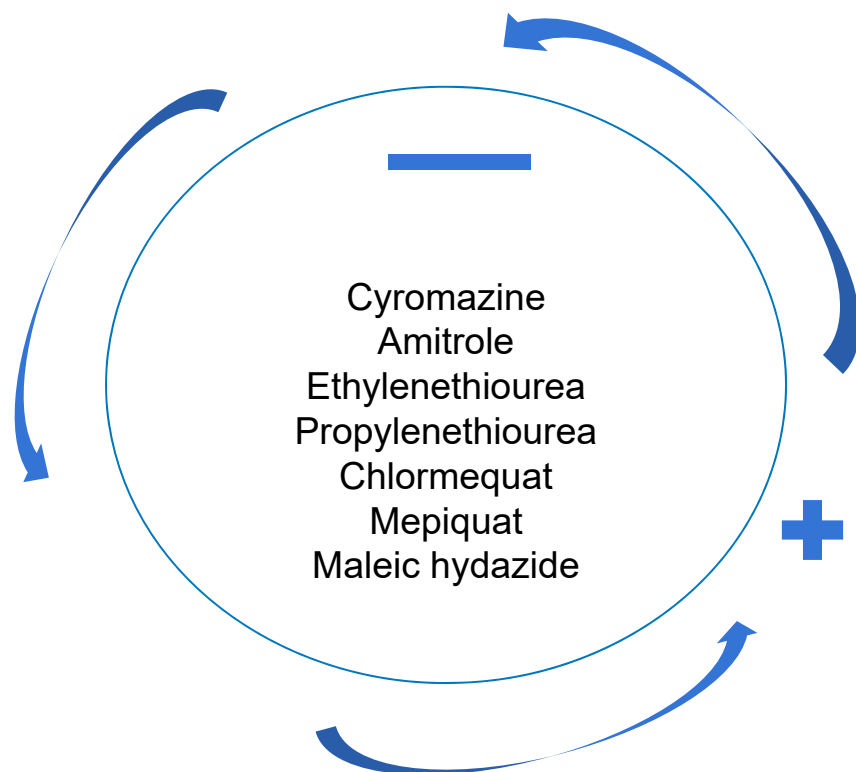
Maximum Residue Limits (**MRLs**), are legislative limits on the concentrations of residual pesticides and their metabolites in fresh food.

**Glyphosate** used as a desiccant on cereal crops to aid harvest- results in a increased frequency of residues in cereal-based products such as bread and breakfast cereals and beer.

**Ethephon** – approved but frequent MRL violations 2016 figures from RASFF, 4 alerts and 3 information for attention notifications covering, grapes, tomatoes, peppers and figs

**Chlorate** – Biocide banned by EU in 2010 because of health risks  
**Perchlorate** – EU established MRLS of 10 µg/kg for most foods in 2015

## Ionic polar pesticides



## Options for Analysis and Sample preparation

### Sample preparation

Derivatization with FMOC?

Extraction with aqueous –  
Narong's et al. method

Extraction organic solvents –  
EURL Quppe



### Liquid Chromatography

Reversed-phase (RP) LC

Ion chromatography (IC)

Porous graphitised/graphitic carbon (PGC)

Hydrophilic interaction liquid chromatography  
(HILIC)

Mixed mode



# Derivatization - using FMOc reagent Glyphosate, AMPA and Glufosinate

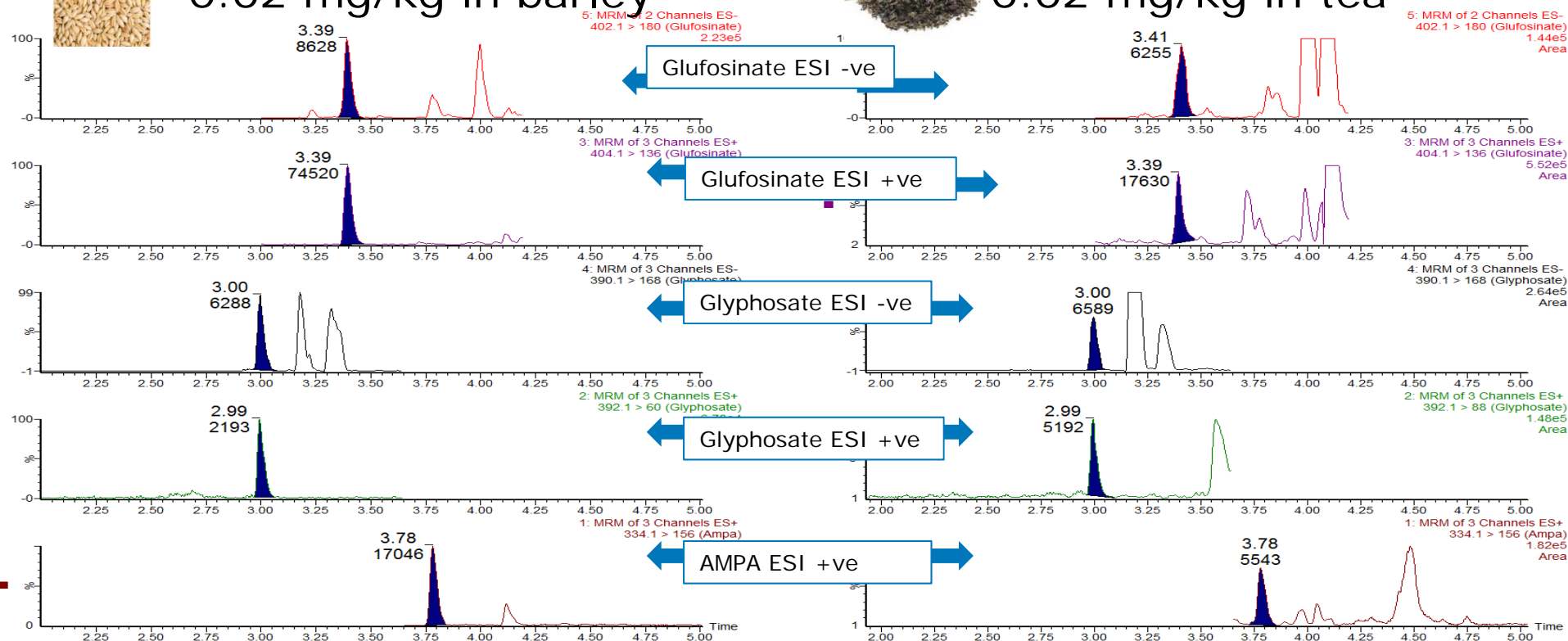
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0.02 mg/kg in barley



0.02 mg/kg in tea



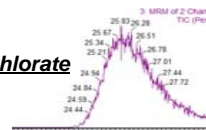
## Our experiences so far ...



### ***HILIC type column (polyvinyl alcohol)***

- + Retention
- +/- Separation of critical pairs
- Ret time stability (> 1min)
- Perchlorate compatibility (5min wide peak)

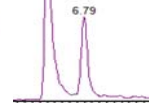
#### ***Perchlorate***



#### ***Separation***

##### ***AMPA***

##### ***Fos AI***



#### ***Retention time stability*** ***Glyphosate***

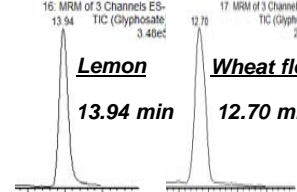
16: MRM of 3 Channels ES-TIC (Glyphosate) 3.40sec  
17: MRM of 3 Channels ES-TIC (Glyphosate) 2.29sec

##### ***Lemon***

13.94 min

##### ***Wheat flour***

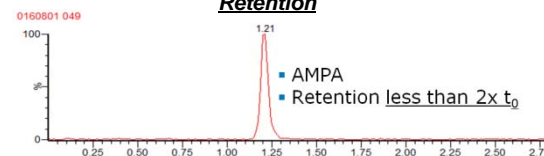
12.70 min



### ***Mixed mode column***

- Retention
- Separation of critical pairs
- Ret time stability

#### ***Retention***

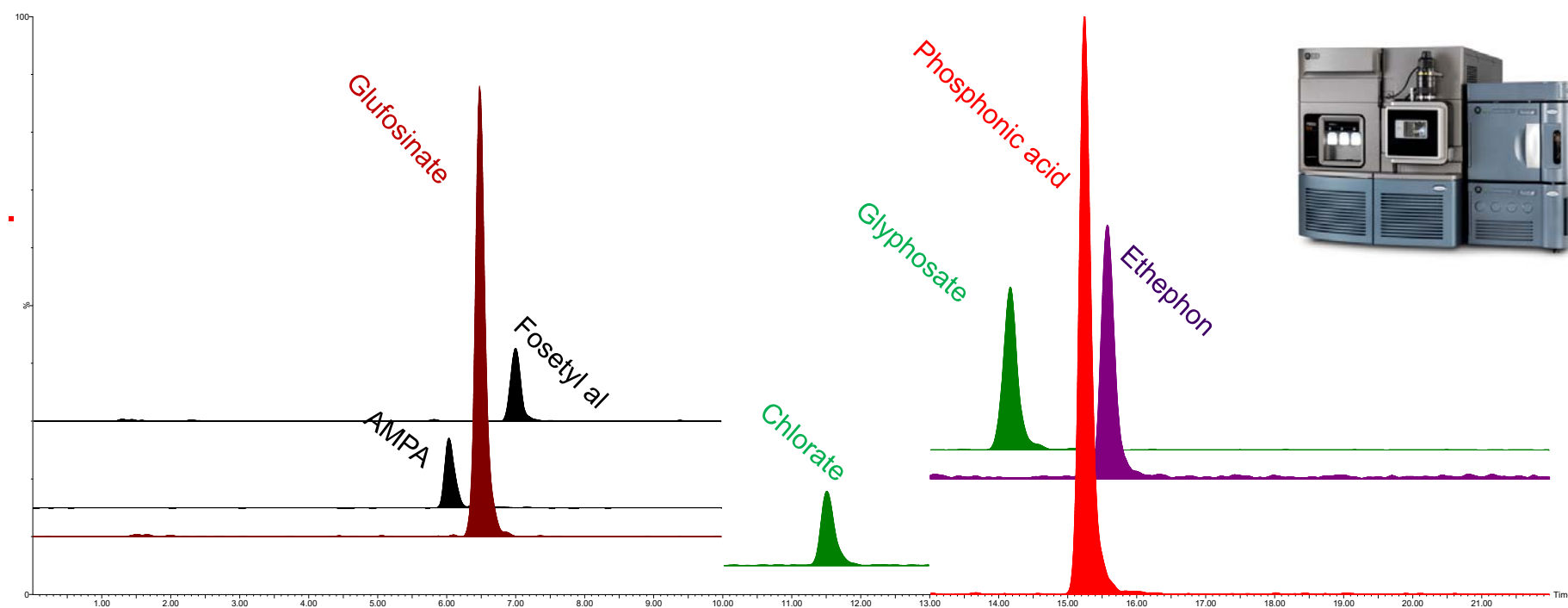


***RSDs > 20% in different commodities***

## Chromatographic separation: HILIC Polyvinyl alcohol based LC column

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- 0.01 mg/kg in beer and extracted as per the EURL Quick Polar Pesticides Extraction method
- MP A: 68:12: 20 water: 45mM ammonium bicarbonate: MeCN; MP B: 50mM ammonium bicarbonate





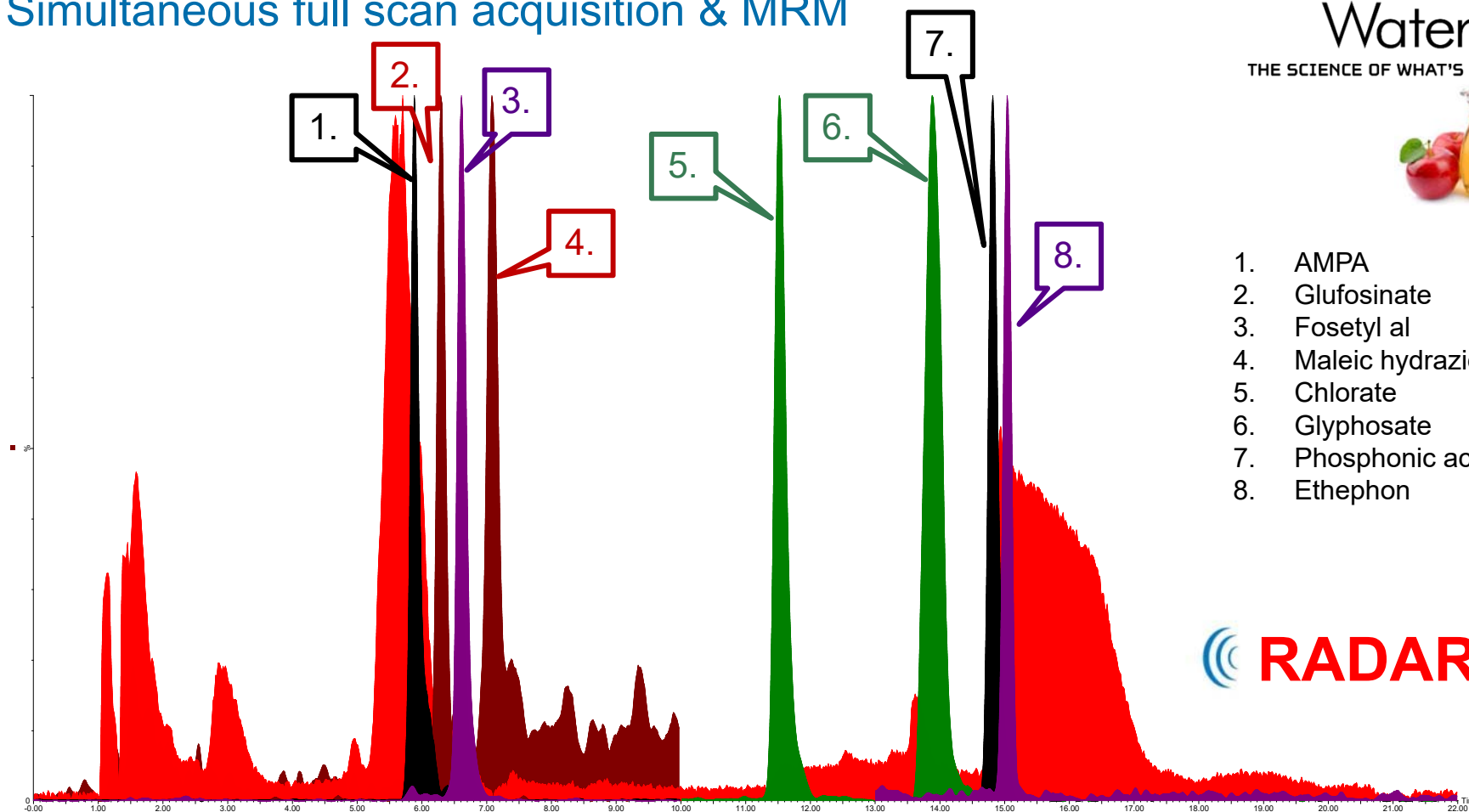
## Simultaneous full scan acquisition & MRM

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1. AMPA
2. Glufosinate
3. Fosetyl al
4. Maleic hydrazide
5. Chlorate
6. Glyphosate
7. Phosphonic acid
8. Ethephon

(( RADAR ))



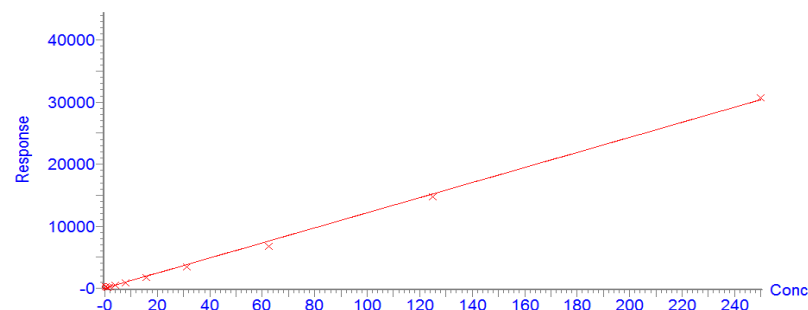
# Quantitation by LC-MS/MS: Glyphosate; 1 to 250 ppb

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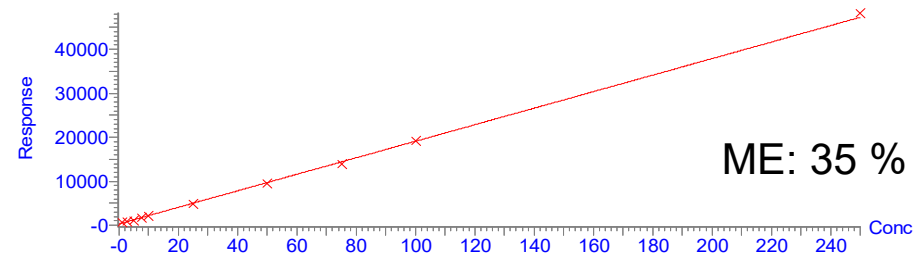
Compound name: Glyphosate  
Correlation coefficient:  $r = 0.999013$ ,  $r^2 = 0.998027$   
Calibration curve:  $121.528 * x + 18.5267$   
Response type: External Std, Area  
Curve type: Linear, Origin: Exclude, Weighting: 1/x, Axis trans: None

Solvent standard



Compound name: Glyphosate  
Correlation coefficient:  $r = 0.999543$ ,  $r^2 = 0.999087$   
Calibration curve:  $187.972 * x + 303.684$   
Response type: External Std, Area  
Curve type: Linear, Origin: Exclude, Weighting: 1/x, Axis trans: None

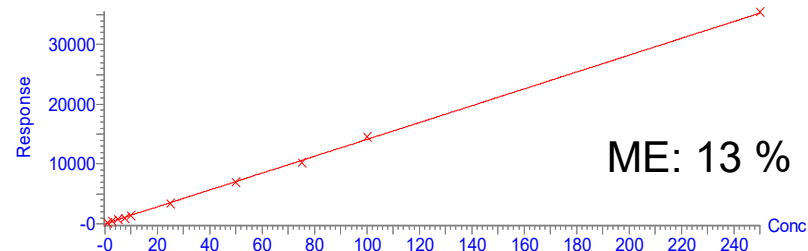
Apple juice



ME: 35 %

Compound name: Glyphosate  
Correlation coefficient:  $r = 0.999464$ ,  $r^2 = 0.998929$   
Calibration curve:  $140.983 * x + 32.7736$   
Response type: External Std, Area  
Curve type: Linear, Origin: Exclude, Weighting: 1/x, Axis trans: None

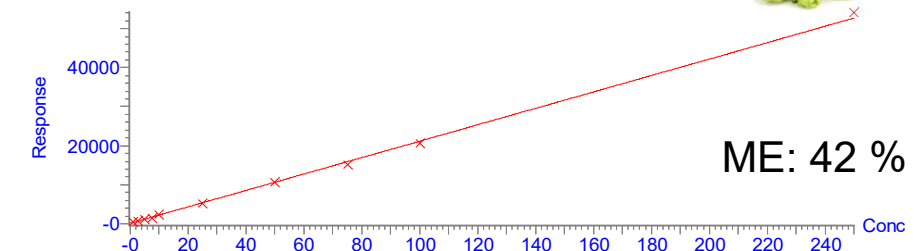
Tomato juice



ME: 13 %

Compound name: Glyphosate  
Correlation coefficient:  $r = 0.999286$ ,  $r^2 = 0.998572$   
Calibration curve:  $210.302 * x + 132.7$   
Response type: External Std, Area  
Curve type: Linear, Origin: Exclude, Weighting: 1/x, Axis trans: None

Beer



ME: 42 %

## Recoveries and repeatability

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			Apple juice		Tomato juice		Beer	
	Conc (mg/kg)	n	Mean (mg/kg)	RSD (%)	Mean (mg/kg)	RSD (%)	Mean (mg/kg)	RSD (%)
Glufosinate	0.01	9	0.0092	9.8	0.0100	9.0	0.0099	5.7
	0.05	9	0.0501	4.9	0.0521	3.9	0.0528	4.0
	0.10	9	0.1039	4.6	0.0980	2.9	0.1047	3.6
Glyphosate	0.01	9	0.0099	8.5	0.0106	9.8	0.0107	4.5
	0.05	9	0.0507	6.1	0.0508	3.8	0.0549	5.7
	0.10	9	0.1046	6.0	0.0961	2.0	0.1068	3.1
Ethephon	0.01	9	0.0095	8.7	0.0097	7.7	0.0106	6.2
	0.05	9	0.0457	6.3	0.0522	3.4	0.0541	5.7
	0.10	9	0.0934	5.0	0.1006	3.5	0.1055	3.7
AMPA	0.01	9	0.0108	9.8	0.0094	2.1	0.0100	7.4
	0.05	9	0.0498	9.0	0.0460	5.4	0.0542	5.6
	0.10	9	0.1011	8.1	0.0912	2.8	0.1060	5.1
Fosetyl AI	0.01	9	0.0095	9.0	0.0090	6.6	0.0106	4.4
	0.05	9	0.0518	4.5	0.0440	1.6	0.0548	5.2
	0.10	9	0.1061	3.2	0.0900	1.5	0.1054	4.3

## Our experiences so far ...

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**Torus Technology**

**HILIC type column**

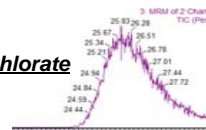
- + Retention
- + Separation of critical pairs
- + Ret time stability
- + Perchlorate compatibility



**HILIC type column (polyvinyl alcohol)**

- + Retention
- +/- Separation of critical pairs
- Ret time stability (> 1min)
- Perchlorate compatibility (5min wide peak)

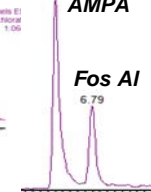
**Perchlorate**



**Separation**

**AMPA**

**Fos AI**



**Retention time stability  
Glyphosate**

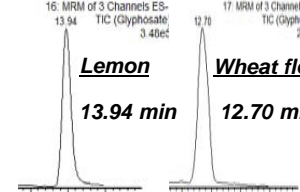
16. MRM of 3 Channels ES- TIC (Glyphosate) 3.40sec  
17. MRM of 3 Channels ES- TIC (Glyphosate) 2.29sec

**Lemon**

**13.94 min**

**Wheat flour**

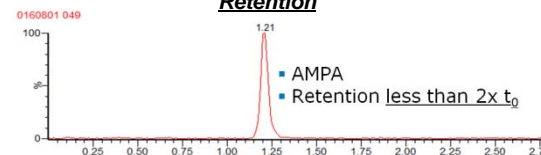
**12.70 min**



**Mixed mode column**

- Retention
- Separation of critical pairs
- Ret time stability

**Retention**



**RSDs > 20% in different commodities**

## Calculating Column 'Dead' Volume $t_0$

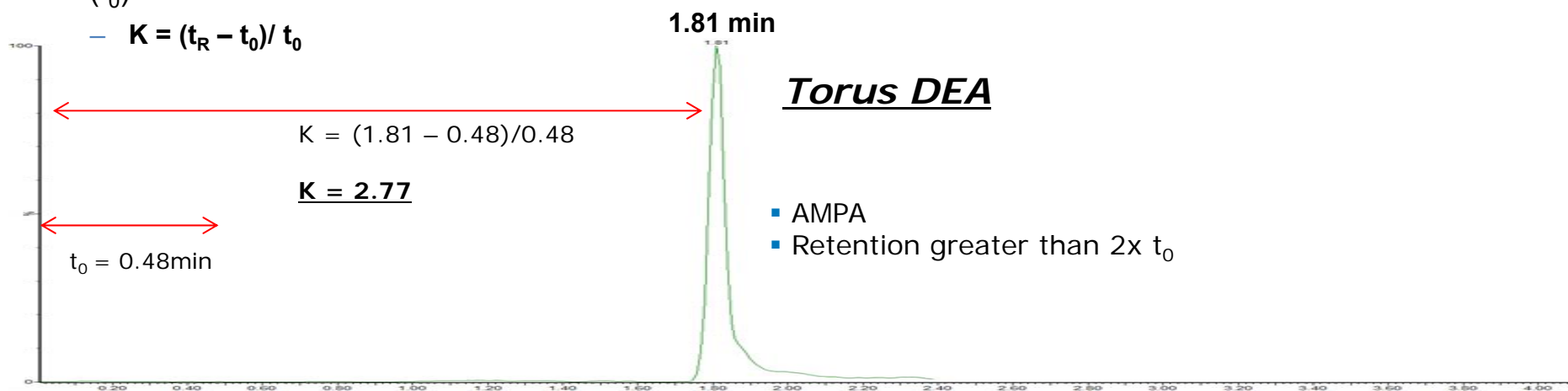
### TORUS DEA 2.1 x 100mm

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- Time required for one volume of mobilephase to exit the column
  - **No retention occurs before the  $t_0$**
- $t_0$  is needed to calculate retention factor
  - $t_0 = V/F$
- **Retention Factor (K)** is the measurement of a columns retention in relation to the column 'dead volume' ( $t_0$ )
  - $K = (t_R - t_0) / t_0$



2.1mm x 100mm  
 $V = 0.24\text{ml}$   
 $F = 0.50\text{ml/min}$



## Chromatography: Example in spinach at 0.01 mg/kg

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Perchlorate  $m/z$  99>83



HEPA  $m/z$  125>79



$m/z$  110>63

AMPA



$m/z$  109>81

Fosetyl AI



MPPA

$m/z$  151>125



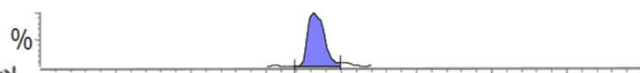
Ethephon

$m/z$  143>107



Glufosinate

$m/z$  180>85



NAG

$m/z$  222>136



Chlorate

$m/z$  83>67

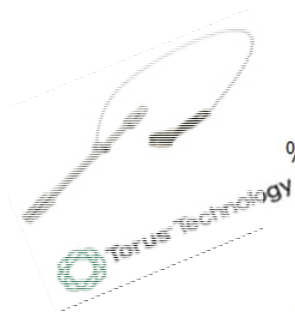


Glyphosate

$m/z$  168>63

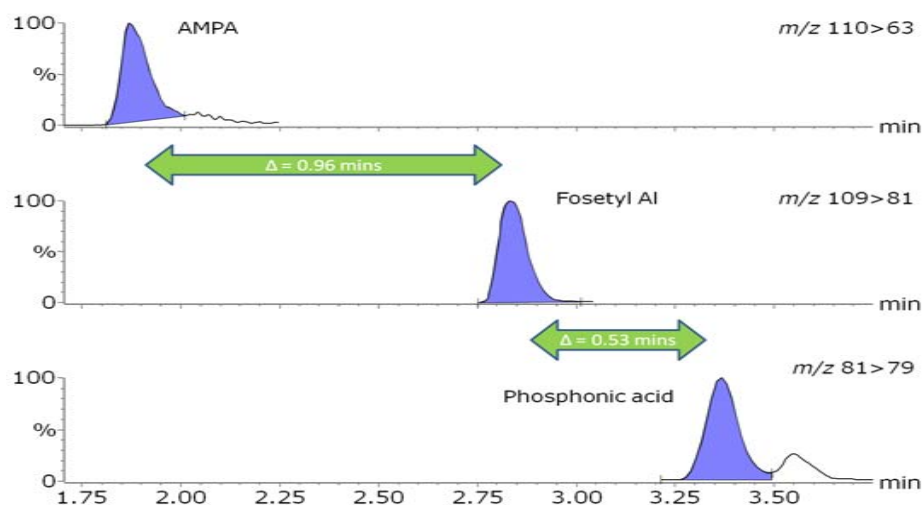


✓ All retained

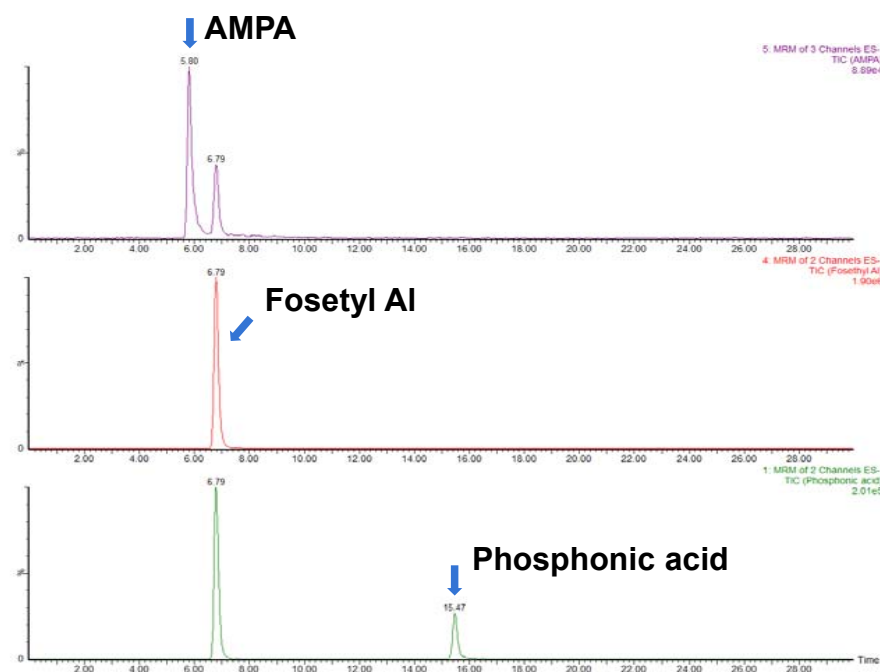


# Chromatography: Critical compounds

## TORUS column



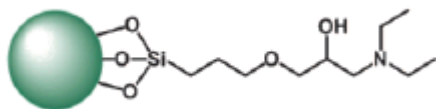
## HILIC type column (polyvinyl alcohol)



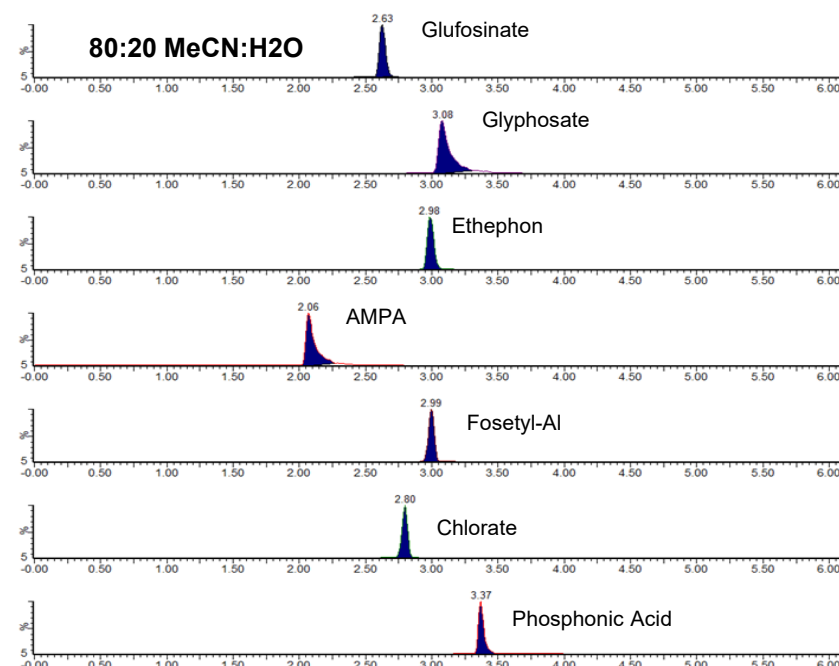
## Torus DEA – Polar pesticides analysis

### Some background

- **Torus DEA, 130Å, 1.7µm, 2.1 mm x 100 mm**
- DEA: Diethylamine - HILIC/WAX
- Ethylene bridged hybrid (BEH) particle
- Two stage functionalization
  - Control retention characteristics
  - Selectivity and peak shape



Patented Approved Technology



(methodology patent pending)



## Experimental: MS parameters



Capillary voltage (kV)	2.4
Source temperature (°C)	150
Desolvation temperature (°C)	600
Cone gas flow (L/Hr)	300
Desolvation gas flow (L/Hr)	1000
Nebuliser (bar)	7

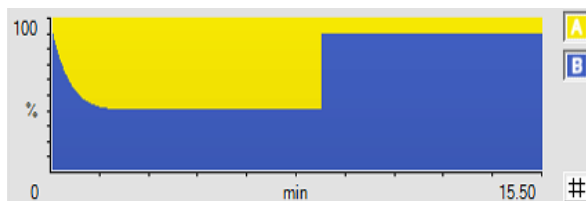
## Waters

<u>Compound</u>	<u>Ion mode</u>	<u>Transitions</u>	<u>Cone voltage (V)</u>	<u>Collision energy (eV)</u>
Glyphosate	ESI-	<b>167.85 &gt; 62.80</b> 167.85 > 80.80 167.85 > 149.85	15	15 15 10
AMPA	ESI-	<b>109.80 &gt; 62.80</b> 109.80 > 78.80 109.80 > 80.80	15	15 15 10
Glufosinate	ESI-	179.90 > 62.80 <b>179.90 &gt; 84.85</b> 179.90 > 134.00	15	25 17 16
Chlorate	ESI-	<b>82.85 &gt; 66.80</b> 82.85 > 50.80	15	14 15
Ethephon	ESI-	142.85 > 78.75 <b>142.85 &gt; 106.85</b>	15	13 8
Fosetyl Aluminium	ESI-	108.85 > 62.80 <b>108.85 &gt; 80.80</b>	15	16 10
Phosphonic acid	ESI-	80.80 > 62.80 <b>80.80 &gt; 78.80</b>	15	15 10
Perchlorate	ESI-	98.80 > 66.90 <b>98.80 &gt; 82.85</b>	20	45 18
Ethephon Hydroxy	ESI-	<b>124.80 &gt; 78.80</b> 124.80 > 94.80 124.80 > 106.90	15	14 12 11
MPPA	ESI-	150.70 > 62.80 150.70 > 106.85 <b>150.70 &gt; 132.85</b>	15	25 16 12
N-Acetyl-Glufosinate	ESI-	221.90 > 58.90 <b>221.90 &gt; 135.90</b> 221.90 > 161.90	20	14 20 12

## Experimental: LC conditions

<b>Column</b>	Torus DEA column (130Å, 1.7 µm, 2.1 x 100 mm)
<b>LC System</b>	I Class FL
<b>Solvent A</b>	50 mM Ammonium Formate pH 2.9 (0.9% Formic Acid)
<b>Solvent B</b>	MeCN + 0.9% Formic Acid
<b>Column Temp</b>	50°C
<b>Sample Temp</b>	10°C
<b>Injection Volume</b>	10 µL
<b>Flow rate</b>	0.5 mL/min

Time (min)	%A	%B	Curve
0	10	90	-
4.50	60	40	2
8.50	60	40	6
15.50	10	90	1



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(methodology patent pending)

## Experimental: Quick Polar Pesticides (QuPPe) extraction procedure

Weighed homogenized sample (5g) into 50 ml tube

Added methanol (10 ml) containing 1 % formic acid

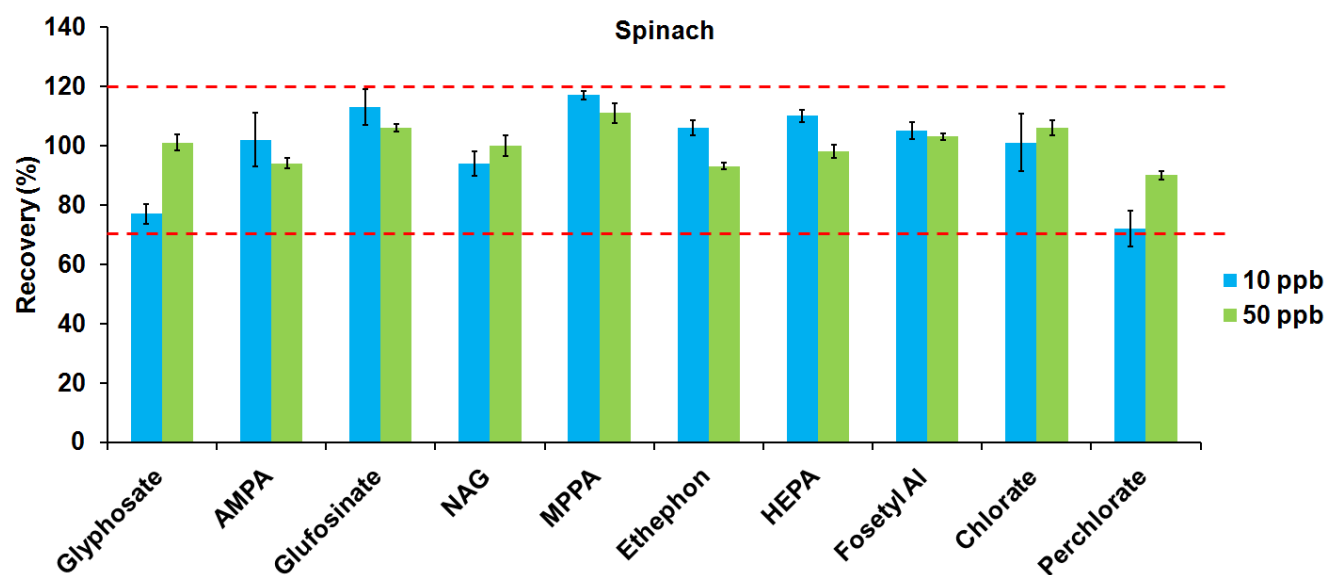
Vortexed thoroughly for 2 minutes

Centrifuged at 6000 rpm for 5 minutes

Filtered supernatant (0.25µm, PVDF, spin filters) into a plastic vial

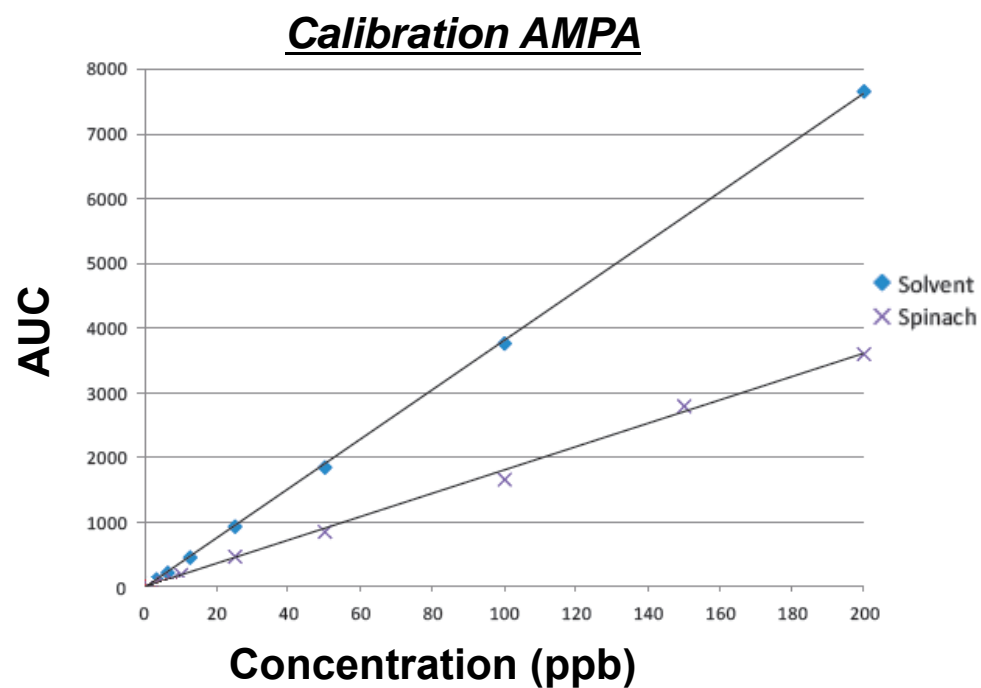


## Method performance: Recoveries and repeatability

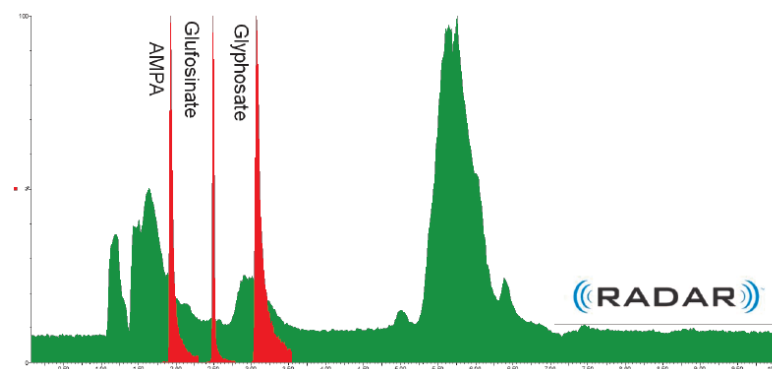


Incurred residue of phosphonic acid was detected and so has been omitted from the results

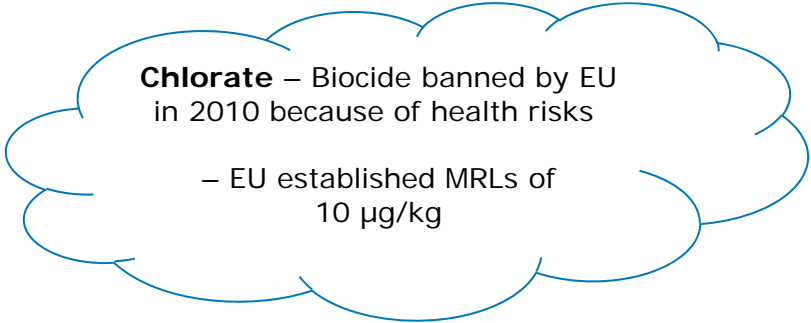
## Matrix effects



	Spinach (%)	
N-Acetyl-Glufosinate	233	Enhancement
Glufosinate	99	
Glyphosate	104	
MPPA	488	
Ethephon	287	
Fosetyl AI	107	No effect
AMPA	47	
Perchlorate	494	Suppression
Chlorate	245	
Ethephon hydroxy	156	
Phosphonic acid	151	



## Chlorate and Perchlorate Analysis



**Chlorate** – Biocide banned by EU  
in 2010 because of health risks

– EU established MRLs of  
10 µg/kg

## Update: Perchlorate and Chlorate

### ■ How

- An alternative LC-MS/MS method with chromatographic separation achieved on the TORUS DEA column, applying an ammonium formate mobile phase gradient.

### ■ Benefits

- Short seven minute run time
- UPLC technology providing optimal peak shapes
- Excellent retention and separation, separation is key due to isobaric interference of perchlorate in chlorates transition.
- Excellent linearity and sensitivity



# Torus DEA

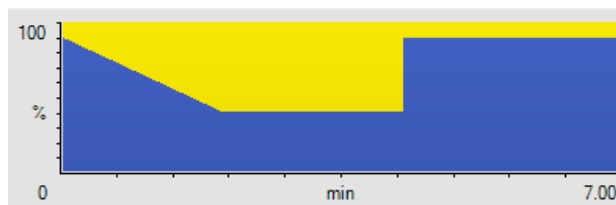
## Chlorate and Perchlorate analysis - LC conditions

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<b>Column</b>	Torus DEA column (130Å, 1.7 µm, 2.1 x 50 mm)
<b>LC System</b>	I Class FL
<b>Solvent A</b>	50 mM Ammonium Formate pH 2.9 (0.9% Formic Acid)
<b>Solvent B</b>	MeCN + 0.9% Formic Acid
<b>Column Temp</b>	50°C
<b>Sample Temp</b>	10°C
<b>Injection Volume</b>	3.0 µL
<b>Flow rate</b>	0.5 mL/min

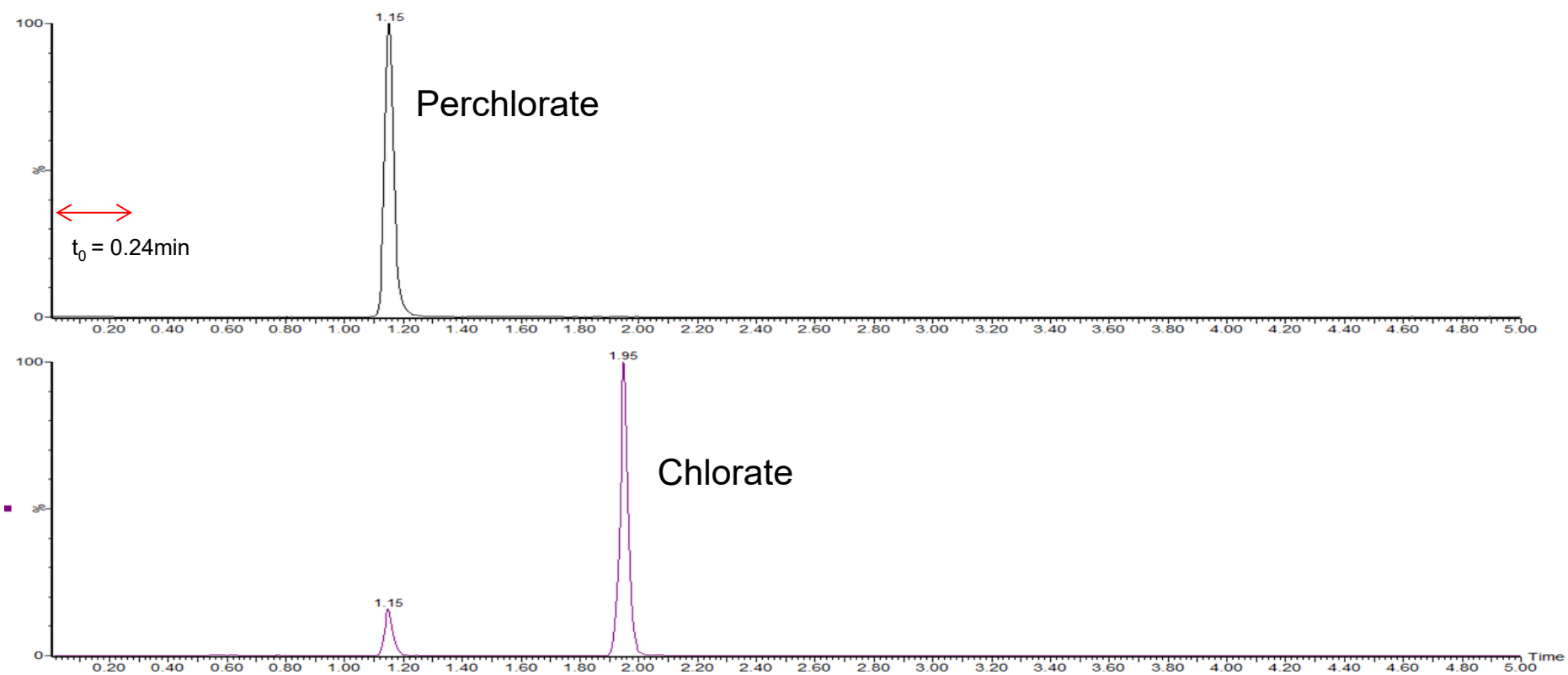


Time (min)	%A	%B	Curve
0	10	90	-
2.00	60	40	6
4.25	60	40	6
7.00	10	90	1





Red Grape:  
0.01 mg/kg spiked post QuPPE extraction

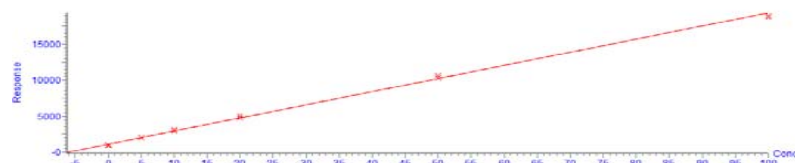


# Grape QuPPE Extraction: Post Spike Matrix Matched 0.010mg/kg – 0.2mg/kg

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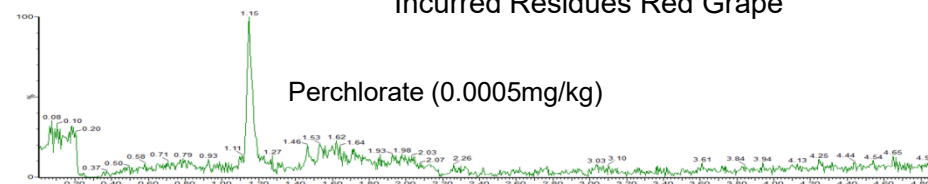
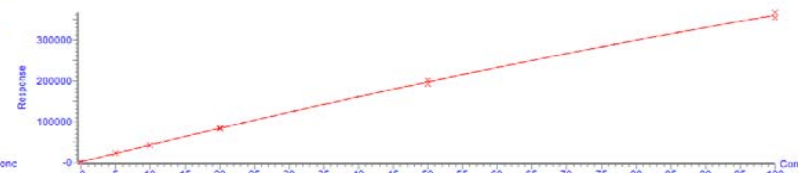


Compound name: Chlorate  
Correlation coefficient:  $r^2 = 0.999027$ ,  $r^2 = 0.997257$   
Calibration curve:  $102.58 \cdot x + 1109.03$   
Response type: External Std. Area  
Curve type: Linear, Origin: Exclude, Weighting: 1/x, Axis trans: None  
Standard Addition Concentration: 0.00075

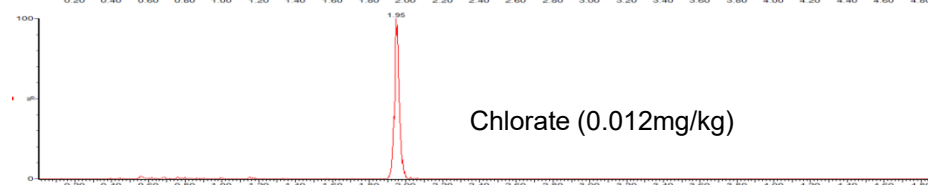


Incurred Residues Red Grape

Compound name: Perchlorate  
Coefficient of Determination:  $R^2 = 0.999272$   
Calibration curve:  $-8.62587 \cdot x^2 + 4247.72 \cdot x + 1195.38$   
Response type: External Std. Area  
Curve type: 2nd Order, Origin: Exclude, Weighting: 1/x, Axis trans: None  
Standard Addition Concentration: 0.201416



Perchlorate (0.0005mg/kg)



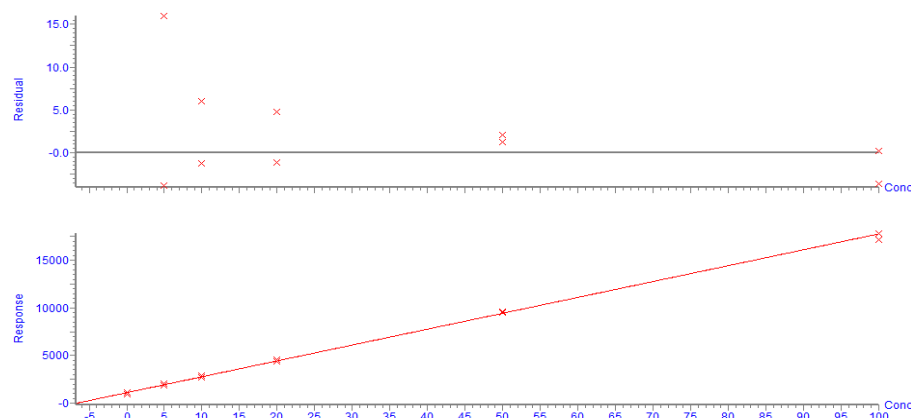
Chlorate (0.012mg/kg)

	Perchlorate %RSD n=6	Chlorate %RSD n=6
0.010mg/kg Post Spike	1.30	2.73*
0.020mg/kg Post Spike	0.96	2.87

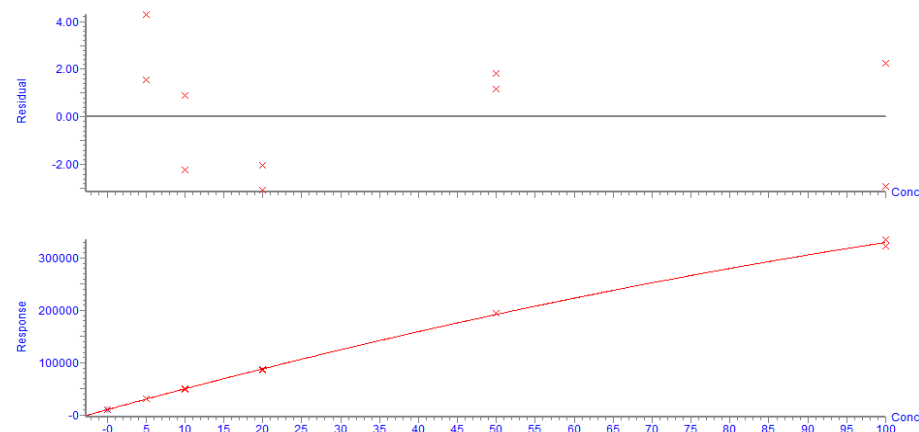
# Cucumber: Spiked post QuPPE extraction

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Compound name: Chlorate  
Correlation coefficient:  $r = 0.999025$ ,  $r^2 = 0.998051$   
Calibration curve:  $165.698 \cdot x + 1101.49$   
Response type: External Std. Area  
Curve type: Linear, Origin: Exclude, Weighting:  $1/x$ , Axis trans: None  
Standard Addition Concentration: 6.60769



Compound name: Perchlorate  
Coefficient of Determination:  $R^2 = 0.999506$   
Calibration curve:  $-8.89849 \cdot x^2 + 4084.44 \cdot x + 10434$   
Response type: External Std. Area  
Curve type: 2nd Order, Origin: Exclude, Weighting:  $1/x$ , Axis trans: None  
Standard Addition Concentration: 2.55458

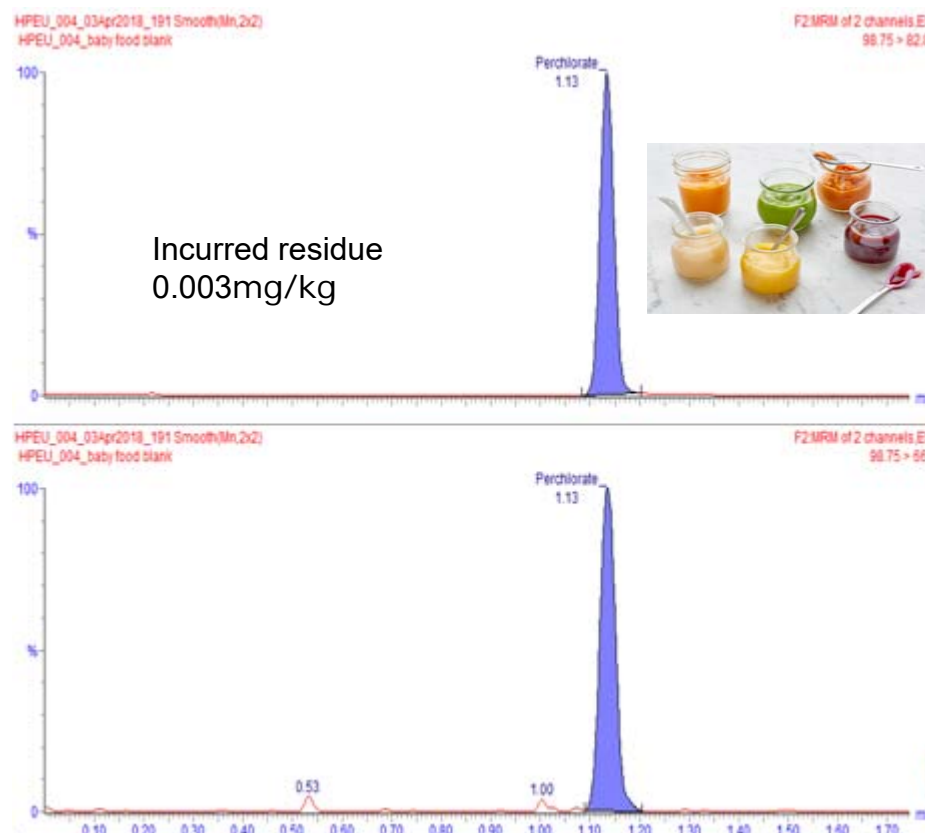
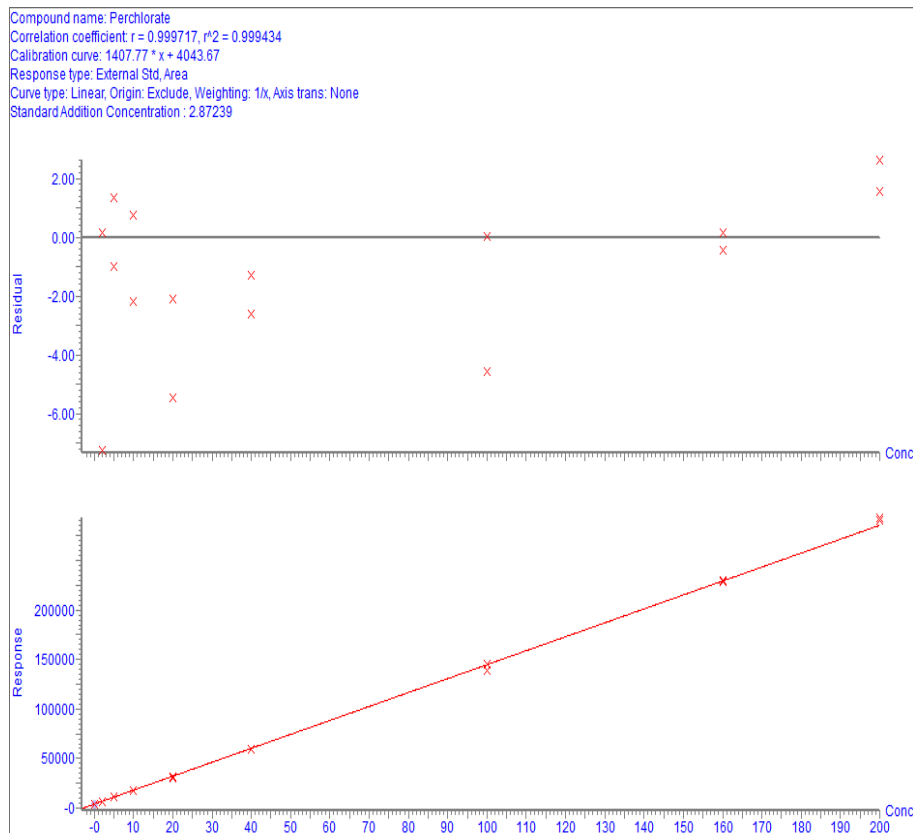


	Perchlorate %RSD n=6	Chlorate %RSD n=6
0.005 mg/kg	0.78	3.28
0.01 mg/kg	1.49	2.63



# Infant Food QuPPE Extraction: Post Spike Matrix Matched 0.002mg/kg – 0.2mg/kg

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## Conclusions

- Expanding on previous LC-MS/MS methods, initial work using the Torus DEA has demonstrated excellent performance for the reliable analysis of polar pesticides in food
- Key benefits include:
  - Improved chromatographic performance for a broad scope of anionic pesticides in a single injection
  - Suitable application for checking compliance with **EU MRLs** and in agreement with **SANTE** guidelines
  - Maintained system sensitivity with LOQs < 0.01 mg/kg in different commodities
  - Repeatable quantitative analysis, with RSDs < 10% achieved at 0.01 mg/kg in spinach without isotopically labelled internal standards
  - Incurred residues of analytes accurately quantified using standard addition calibration

# Acknowledgements

## ■ References

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## ■ Acknowledgements

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- UGRL, Turkey
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- Galab, Germany



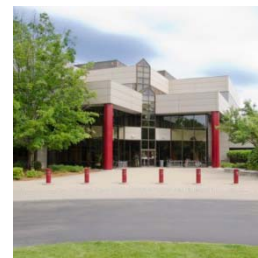
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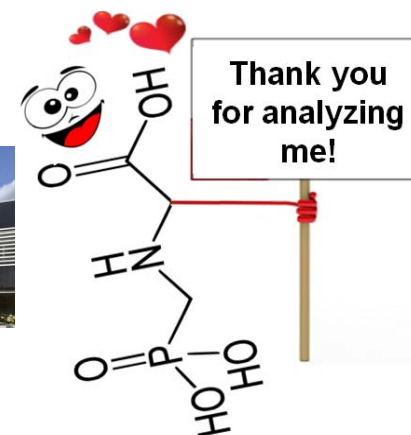


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# Thank you for your attention

**Any questions**



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